CB1 Receptor Antagonist Precipitates Withdrawal in Mice Exposed to $\Delta^9$-Tetrahydrocannabinol

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ABSTRACT
Although the question of cannabinoid dependence had been very controversial until the discovery of a cannabinoid antagonist, SR141716A. The objective of this study was to develop and characterize a mouse model of precipitated withdrawal indicative of cannabinoid dependence. Using a dosing regimen known to produce pharmacological and behavioral tolerance, mice were treated with $\Delta^9$-tetrahydrocannabinol ($\Delta^9$-THC) twice a day for 1 wk. SR141716A administration after the last $\Delta^9$-THC injection promptly precipitated a profound withdrawal syndrome. Typical withdrawal behavior was an increase in paw tremors and head shakes that was accompanied with a decrease in normal behavior such as grooming and scratching. Of the three $\Delta^9$-THC regimens tested, daily $\Delta^9$-THC injections of 10 and 30 mg/kg produced the greatest number of paw tremors and head shakes and the least number of grooms after challenge with SR141716A. Precipitated withdrawal was apparent after 2, 3, 7 and 14 days of treatment based on an increase in paw tremors in $\Delta^9$-THC-treated mice as compared with vehicle-treated mice. These findings are consistent with SR141716A-precipitated withdrawal in rats. Moreover, these results suggest that mice are a viable model for investigating dependence to cannabinoids.

With the extensive use of marijuana recreationally and its promotion as a therapeutic agent for the treatment of emesis, pain, and loss of appetite, the adverse consequences of chronic exposure become increasingly important. As a result, tolerance and dependence to cannabinoids have generated renewed interest. Tolerance is known to develop to most of the pharmacological effects of $\Delta^9$-THC that included antinociception, hypothermia, depression of locomotor activity, catalepsy, anticonvulsant activity, ataxia, hypotension, immunosuppression and schedule-controlled behavior (Compton et al., 1990; Kaymakcalan, 1973; McMillan et al., 1971; Pertwee, 1991). As with $\Delta^9$-THC, the pharmacological effects of other psychoactive cannabinoids, especially CP-55,940, have also been shown to undergo tolerance development (Fan et al., 1994; Pertwee et al., 1993; Pertwee, 1991).

Dependence and tolerance often develop concomitantly, and in some instances, the severity of the withdrawal syndrome is a function of the magnitude of tolerance development. Thus based on the tolerance data for cannabinoids, one would predict that dependence could develop to cannabinoids. An abstinence syndrome has been described in humans after cessation of chronic marijuana treatment (Jones, 1983). However, some of the limitations of the human studies included lack of placebo and double-blind controls, confinement of individuals to a hospital for long periods of time (20-30 days) and knowledge of drug treatment which may have contributed to anticipation of subjective withdrawal effects; such as dysphoria; upon drug cessation.

Several studies of $\Delta^9$-THC abstinence in nonhuman animals have been carried out. In monkeys chronically administered $\Delta^9$-THC, abstinence produced tremors, twitching, aggression, anorexia, hyperirritability and disruption of schedule-controlled behavior (Beardsley et al., 1986; Kaymakcalan, 1979). Readmission of $\Delta^9$-THC reversed the disruption in the schedule-controlled behavior, but it was unclear if $\Delta^9$-THC readministration reversed the other effects. Similarly, pigeons given daily i.m. injections of very high doses of $\Delta^9$-THC displayed a disruption in schedule-controlled behavior shortly after drug cessation (McMillan et al., 1971). However, these investigators were unable to reverse this decrement in performance by drug readministration. Other researchers have reported that cessation of chronic $\Delta^9$-THC treatment resulted in an increase in grooming behavior and motor activity in rats (Kaymakcalan et al., 1977; Sjoden, 1973). However, this observation has not been observed consistently by other laboratories. One group of

ABBREVIATIONS: $\Delta^9$-THC, $\Delta^9$-tetrahydrocannabinol; SR141716A, N-(piperidin-1-yl)-5-(4-chlorophenyl)-1-(2,4-dichlorophenyl)-4-methyl-1H-pyrazole-3-carboxyamide; C.L., confidence limits.
investigators found withdrawal signs only when chronically treated Δ9-THC rats were challenged with a high dose of naloxone (Hirschhorn and Rosecrans, 1974). Others reported an abstinent syndrome that occurred only after the administration of neurotransmitter reuptake inhibitors, either clozapine, imipramine or fluoxetine, the day after cessation of chronic Δ9-THC treatment (Taylor and Fennessy, 1982; Verberne et al., 1980). It is unclear whether these behavioral effects were due to withdrawal or a drug interaction.

An effective means of demonstrating dependence is to precipitate physical withdrawal by challenging chronically treated animals with an appropriate antagonist. Discovery of a long-awaited cannabinoid antagonist has made it possible to assess dependence by conducting precipitated withdrawal studies. The novel competitive cannabinoid antagonist SR141716A binds with high affinity to the central CB1 cannabinoid receptor and effectively antagonizes a variety of cannabinoid effects in rodents (Compton et al., 1996; Rinaldi-Carmona et al., 1994, 1996). Recently, it was shown that SR141716A was capable of precipitating a withdrawal syndrome in rats chronically treated with Δ9-THC (Aceto et al., 1995, 1996; Tsou et al., 1995). Upon termination of Δ9-THC treatment and immediate administration of SR141716A, a profound abstinence syndrome was evident by the appearance of overt behavioral signs in these chronic drug-treated rats. The withdrawal signs included wet-dog shakes, involuntary paw tremors, ptosis, tongue rolling, retropulsion, head shakes and facial rubbing. These studies were the first to demonstrate unequivocally that chronic cannabinoid treatment resulted in a physical withdrawal syndrome in rats.

An important question is whether precipitated cannabinoid withdrawal occurs in other species. Our knowledge regarding the pharmacological actions of cannabinoids has been derived from numerous animal species. For example, pharmacological tolerance and cross-tolerance studies with Δ9-THC, CP-55,940, WIN 55-212 and anandamide have been established using mouse behavioral models and smooth muscle preparations (Fan et al., 1994; Pertwee, 1993, Pertwee et al., 1992, 1995). In addition, localization of the cannabinoid receptor and second messenger systems and changes in receptor number and mRNA levels for the cannabinoid receptor have been investigated in mice (Abood et al., 1993; Fan et al., 1996; Herkenham et al., 1991). In short, development of cannabinoid dependence in a second species, such as the mouse, would provide further credence for this phenomenon that has been characterized in the rat. Additionally, the development of a mouse model of dependence would be beneficial since so much is known regarding the cannabinoid system in this species.

**Materials and Methods**

**Animals.** Male ICR mice (Harlan Laboratories, Dublin, VA) weighing 20 to 27 g and housed six mice per cage were used in all experiments. Mice were maintained on a 14:10 hr light:dark cycle with water and food *ad libitum.*

**Drug preparation.** Δ9-THC was provided by the National Institute on Drug Abuse, Rockville, MD, and SR141716A was generously donated by Pfizer Central Research (Groton, CT). All drugs were dissolved in a 1:1:18 solution of ethanol, emulphor and 0.9% saline. Emulphor (EL-620, a polyoxyethylated vegetable oil, GAF Corporation, Linden, NJ) is currently available as Alkumulphor. All s.c. and i.p. injections were administered at a volume of 0.1 ml/10 g of body weight. On test day mice were acclimated in the laboratory overnight without interruption of food and water.

**Tolerance development and antagonist challenge.** For six days either Δ9-THC (10 mg/kg) or vehicle (1:1:18) was given s.c. twice a day, once between 09.00-11.00 hr and again between 21.00-23.00 hr. This regimen has been shown to produce tolerance to the antinoceptive, locomotor, hypothermic and cataleptic effects of Δ9-THC (Abood et al., 1993; Fan et al., 1994). Body weights were recorded and used as an indicator of toxicity. On day 7, the test day, mice received an acute i.p. challenge with either SR141716A or vehicle 4 hr after their last chronic Δ9-THC treatment. For studying the time course of dependence development, the same protocol was used except in addition to a 7-day dosing regimen, separate groups of mice were also dosed for either 1, 2, 3 or 14 days. For example, mice received Δ9-THC (10 mg/kg) or vehicle in the morning followed by SR141716A or vehicle 4 hr later for the day 1 time point.

**Behavioral evaluation.** Immediately after either SR141716A or vehicle challenge, mice were observed for 30 min (except where otherwise noted) in clear activity cages for typical withdrawal behaviors and any unique behavior. These typical behaviors included head shakes (turning or twisting head side to side), paw tremors, retropulsion (more than three steps backward), writhing, scratching, rubbing, grooming, piloerection, penile erection and Straub tail. Paw tremors were rapid lateral movements of the paws that typically lasted several seconds and were episodic which allowed for quantitation. A grooming episode was typically characterized by the licking of paws and body and by rubbing paws over nose, head and ears.

Different mice were used for each test and time point; and there were at least six mice per group. Experimenters were blind to the drug conditions in all experiments. All studies were approved by the Institutional Animal Care and Use Committee.

**Statistics.** Data were analyzed by ANOVA at P < .05. Bonferroni post hoc analyses (comparison with vehicle) were used when appropriate. For dose-response curves, E<sub>max</sub> values were calculated with ALLFIT with the minimum being constrained to vehicle-vehicle values. A modification of the method of Tallarida and Murray (1987) was used to calculate ED<sub>50</sub> values and 95% C.L. ED<sub>50</sub> values were not calculated if one-way ANOVA was not significant at P < .05. Each point represents at least six mice per dose and dose-effect curves consist of at least three doses.

**Results**

Before designing a model of precipitated withdrawal in THC-dependent mice, it was essential to characterize the pharmacological effects of the antagonist alone. Doses of 10 and 30 mg/kg of SR141716A were administered i.p.; and experimenters blind to the drug conditions observed the mice for 30 min for behaviors similar to those reported in rats undergoing precipitated withdrawal, as well as any unique behaviors (fig. 1). The most prominent behavioral signs tallied were paw tremors, head shakes and scratching. There were no significant differences in the number of paw tremors between vehicle and either dose of SR141716A (P = .15). On the contrary, the highest dose of SR141716A, 30 mg/kg, elicited a significant number of head shakes when compared with vehicle and a dose of 10 mg/kg of SR141716A. SR141716A alone elicited a marked increase in scratching in naive mice (P = 14.329, P < .05). Both doses of SR141716A were significantly different from vehicle.

Because the lower dose of SR141716A failed to elicit a significant number of paw tremors and head shakes during the 30-min period after injection, a subsequent experiment was conducted in which mice were challenged with SR141716A (10 mg/kg) and observed for 1 hr. Several behavioral signs were recorded at 15-min intervals to establish the
time course of the acute effects of SR141716A (fig. 2). The data were analyzed by two-way ANOVA for repeated measures. There was no drug effect ($P = .71$) or time effect ($P = .4936$) for paw tremors. SR141716A did, however, elicit a significant drug effect for head shakes ($F = 5.37, P < .05$) but failed to show a time effect ($P = .87$) or interaction ($P = .51$). Because the behavior in mice treated acutely with SR141716A or vehicle did not differ at either 15 or 30 min, or when the data for these two time intervals were collapsed, the dependence experiments were conducted for 30 min immediately after the acute challenge with either of SR141716A (10 mg/kg).

SR141716A markedly increased the number of paw tremors in a dose-dependent fashion in mice chronically treated with $\Delta^9$-THC (fig. 3A). As the dose of SR141716A increased, the number of paw tremors in $\Delta^9$-THC chronically treated mice increased with an $ED_{50}$ value (95% C.L.) of 4.6 mg/kg (2.5-8.2). Mice treated chronically with vehicle and challenged with SR141716A at doses up to 30 mg/kg did not differ significantly from vehicle-vehicle treated mice with respect to paw tremors. Therefore, an $ED_{50}$ value for mice chronically treated with vehicle could not be calculated. SR141716A also dose-dependently increased the number of head shakes in mice, but did so for both groups (fig. 3B). Using ALLFIT analysis and a modification of the method of Tallarida and Murray, the $ED_{50}$ value (95% C.L.) for $\Delta^9$-THC chronically treated mice was 1.3 mg/kg (0.6-2.7). The $ED_{50}$ value for vehicle chronically treated mice could not be calculated because according to ALLFIT, a percent response greater than 25 was never attained.

Upon challenge with SR141716A, the incidence of grooming behavior in mice treated chronically with $\Delta^9$-THC and those treated chronically with vehicle was markedly different (fig. 3C). Vehicle-vehicle treated mice exhibited 6.9 $\pm$ 0.8 (mean $\pm$ S.E.M.) grooming episodes during the 30-min observation period (table 1). A one-way ANOVA indicated that there was no significant effect of SR141716A in mice treated repeatedly with vehicle. However, $\Delta^9$-THC-treated mice showed a dose-dependent decrease in normal grooming behavior when challenged with the antagonist, SR141716A, generating an $IC_{50}$ value of 0.80 mg/kg (0.32-2.0).

Unlike paw tremors and head shakes, the frequency of scratching was dose-responsive only for vehicle-treated mice (fig. 3D). Very little scratching was observed in mice chronically treated with $\Delta^9$-THC (10 mg/kg). A one-way ANOVA was done on each dose-response curve to determine if treatment differed from vehicle control. SR141716A dose-dependently increased scratching in mice treated chronically with vehicle with an $ED_{50}$ value of 4.5 mg/kg (3.7-5.5). Table 1 summarizes the vehicle control mean $\pm$ S.E.M. values for the SR141716A dose-response curves for all four behaviors.

All of the data described above were generated with a chronic treatment regimen of 10 mg/kg of $\Delta^9$-THC. To determine whether the magnitude of the dependence syndrome was dependent upon the $\Delta^9$-THC treatment regimens, separate groups of mice were treated with either 3, 10 or 30 mg/kg of $\Delta^9$-THC or vehicle for 6.5 days and then challenged with an acute dose of either SR141716A (10 or 30 mg/kg) or vehicle on day 7 (fig. 4). The data were analyzed by using two-factor...
ANOVA. Bonferroni post hoc analyses were used when appropriate. A significant Δ9-THC regimen/SR141716A dose interaction resulted for both paw tremors (F\_5 = 7.854, P < .05) and head shakes (F\_5 = 2.446, P < .05). Although there was not a significant Δ9-THC dose/SR141716A dose interaction for grooming behavior (P = .3630), a main effect for Δ9-THC dose existed (F\_5 = 22.96, P < .05). SR141716A did not appear to precipitate physical withdrawal in mice treated chronically with 3 mg/kg of Δ9-THC, because neither dose of SR141716A elicited a significant increase in the number of paw tremors and decrease in grooming behavior as compared with their respective chronic vehicle/SR141716A group. A significant difference also existed for all three behaviors between SR141716A- and vehicle-challenged mice that were treated chronically with a 10-mg/kg dose of Δ9-THC.

Mice treated with 30 mg/kg of Δ9-THC exhibited physical withdrawal signs when challenged with 30 mg/kg of SR141716A. The frequency of paw tremors and head shakes increased although grooming behavior decreased upon administration of 30 mg/kg of SR141716A to the Δ9-THC-treated groups when compared with vehicle/SR141716A (30 mg/kg). However, when the challenge dose was lowered to 10 mg/kg of SR141716A, only grooming behavior exhibited any

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**Table 1**

Vehicle effects after chronic vehicle or Δ9-THC administration

<table>
<thead>
<tr>
<th></th>
<th>Vehicle-vehicle treated</th>
<th>Δ9-THC-vehicle treated</th>
</tr>
</thead>
<tbody>
<tr>
<td>Paw tremors</td>
<td>0.77 ± 0.27</td>
<td>2.0 ± 0.55</td>
</tr>
<tr>
<td>Head shakes</td>
<td>1.0 ± 0.32</td>
<td>0.76 ± 0.27</td>
</tr>
<tr>
<td>Scratching</td>
<td>4.6 ± 1.5</td>
<td>3.4 ± 0.77</td>
</tr>
<tr>
<td>Grooming</td>
<td>6.9 ± 0.84</td>
<td>3.3 ± 0.76</td>
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Mice received either Δ9-THC (10 mg/kg) or vehicle (1:1:18) s.c. twice a day for 6 days and once on day 7. Four hours after the last injection, vehicle was administered i.p. The frequency of paw tremors, head shakes, scratching and grooming behavior was observed for 30 min immediately after the challenge drug or vehicle. Results are presented as mean ± S.E.M. Each point represents 6 to 17 mice.

ANOVA. Bonferroni post hoc analyses were used when appropriate. A significant Δ9-THC regimen/SR141716A dose interaction resulted for both paw tremors (F = 7.854, P < .05) and head shakes (F = 2.446, P = .05). Although there was not a significant Δ9-THC dose/SR141716A dose interaction for grooming behavior (P = .3630), a main effect for Δ9-THC dose existed (F = 22.96, P < .05). SR141716A did not appear to precipitate physical withdrawal in mice treated chronically with 3 mg/kg of Δ9-THC, because neither dose of SR141716A elicited a significant increase in paw tremors in chronic Δ9-THC/SR141716A mice compared to chronic vehicle/SR141716A mice. Δ9-THC (3 mg/kg)/SR141716A did not differ significantly from its respective chronic vehicle/SR141716A group for any of the behaviors reported. The number of paw tremors and head shakes observed in mice chronically treated with a dose of 3 mg/kg of Δ9-THC and challenged with SR141716A significantly differed from those observed in mice chronically treated with a dose of 3 mg/kg of Δ9-THC and challenged with vehicle (fig. 4). Planned comparisons showed no differences in the number of grooms in these mice when challenged with either vehicle or SR141716A. SR141716A markedly induced precipitated withdrawal in mice treated chronically with 10 mg/kg of Δ9-THC as indicated by a significant increase in the number of paw tremors and decrease in grooming behavior as compared with their respective chronic vehicle/SR141716A group. A significant difference also existed for all three behaviors between SR141716A- and vehicle-challenged mice that were treated chronically with a 10-mg/kg dose of Δ9-THC.

Mice treated with 30 mg/kg of Δ9-THC exhibited physical withdrawal signs when challenged with 30 mg/kg of SR141716A. The frequency of paw tremors and head shakes increased although grooming behavior decreased upon administration of 30 mg/kg of SR141716A to the Δ9-THC-treated groups when compared with vehicle/SR141716A (30 mg/kg). However, when the challenge dose was lowered to 10 mg/kg of SR141716A, only grooming behavior exhibited any
difference. Compared with their respective chronic Δ⁹-THC/vehicle group, mice treated with 30 mg/kg of Δ⁹-THC and challenged with 10 mg/kg of SR141716A had significantly more head shakes and less grooming behavior. When challenged with the higher dose of SR141716A (30 mg/kg), mice chronically treated with Δ⁹-THC (30 mg/kg) had significantly more paw tremors and head shakes compared to the chronic Δ⁹-THC/vehicle group as well as the chronic vehicle/SR141716A group. In contrast, the apparent decrease in grooming behavior in these mice was not statistically significant from their respective control groups.

In summary, the existence of precipitated withdrawal in mice treated with Δ⁹-THC for approximately 1 wk was clearly dependent upon the doses of both Δ⁹-THC and SR141716A. To determine the time course for development of dependence, mice were treated with vehicle or Δ⁹-THC for either 1, 2, 3, 7 or 14 days and then challenged with SR141716A (fig. 5). A two-way factorial ANOVA revealed a drug effect (F = 72.782, P < .05), time effect (F = 4.757, P = .05), and interaction between number of days and challenge dose (F = 5.840, P = .05), with respect to paw tremors. The number of paw tremors increased as mice were exposed to a dose of 10 mg/kg of Δ⁹-THC for longer periods of time. Mice treated with Δ⁹-THC for either 2, 3, 7 or 14 days showed a significant increase in paw tremors compared with their respective vehicle group. The incidence of paw tremors was also greater in Δ⁹-THC-treated mice after 2, 3, 7 and 14 days compared with 1 day of Δ⁹-THC treatment. No main time effect (P = .3898) or interaction (P = .705) for head shakes was found in mice treated with Δ⁹-THC. There was however, a main drug effect (F = 22.738, P < .05) with respect to head shakes.

**Discussion**

Usually drug dependence can be established either by abruptly terminating chronic treatment and observing a spontaneous withdrawal syndrome or by precipitating a withdrawal syndrome in chronically treated animals with an appropriate antagonist. Abrupt withdrawal commonly occurs with drugs that do not have a long duration of action. It is reasonable to speculate that spontaneous withdrawal occurs if the biologically active levels of the drug dissipate before the endogenous system can fully recover from the dependent state. Not surprisingly, it is more difficult to detect spontaneous withdrawal with drugs, such as the cannabinoids, that have a long duration of action. Of course, the actions of both long- and short-acting drugs can be abruptly terminated with an antagonist challenge.

Paw tremors appear to be a reliable indicator of cannabinoid dependence. In the present study, paw tremors were the most prominent and dose-responsive withdrawal sign observed. Others have reported involuntary paw tremors and twitching during SR141716A-precipitated withdrawal studies with rats (Tsou et al., 1995) as well as during abrupt withdrawal studies with rats and monkeys (Compton et al., 1990; Kaymakcalan, 1973). Head shakes were another common SR141716A-precipitated withdrawal sign observed in mice and appeared to be consistent with wet-dog shakes in rats that involve both head and body movements (Aceto et al., 1995, 1996; Tsou et al., 1995).

In abrupt and precipitated withdrawal studies of chronic cannabinoids, monkeys and rats exhibited an increase in motor activity (Beardsley et al., 1986; Kaymakcalan et al., 1977; Pertwee, 1991; Tsou et al., 1995) and excessive grooming (Kaymakcalan et al., 1977; Pertwee, 1991). Even though mice showed a remarkable decrease in grooming behavior and scratching during SR141716A-precipitated withdrawal, they still displayed a constantly changing disorganized sequence of movements very similar to that seen in rats during precipitated withdrawal (Tsou et al., 1995). The decrease in normal behavior in cannabinoid-dependent mice was most likely due to the fact that these mice were overwhelmingly preoccupied with paw tremors and head shakes. Paw tremors and other mouse behaviors, such as grooming, are mutually exclusive. Therefore, the mouse model of cannabinoid dependence resembles and confirms the rat model of cannabinoid precipitated withdrawal while adding credibility to the rat and monkey data from abrupt cannabinoid withdrawal studies.

Although mice and rats chronically exposed to Δ⁹-THC and then challenged with SR141716A elicited some common withdrawal signs, it is important to point out that scoring SR141716A-precipitated withdrawal was complicated by the fact that not all animals exhibit identical withdrawal behaviors. Some mice produced primarily paw tremors although head shakes dominated in others. A few mice exhibited little effect other than writhing. Even though strong similarities existed between rats and mice during SR141716A-precipitated withdrawal, differences among the withdrawal syndromes must also be considered. Observation of retropulsion...
and writhing was sporadic among individual mice which was in contrast to the more frequent observation of these behaviors in rats during SR141716A-precipitated withdrawal (Aceto et al., 1996; Tsou et al., 1995). Aceto et al. (1995, 1996) reported a marked appearance of eyelid ptosis and significant increase in facial rubbing in rats during SR141716A-precipitated withdrawal. However, there was not a clear trend in the incidence of eyelid ptosis and facial rubbing in mice.

SR141716A alone had very little effect on naive mice with the exception of increased scratching. It significantly increased scratching in naive mice and dose-dependently increased scratching in vehicle-treated mice. Interestingly, scratching was suppressed in Δ²-THC-treated mice challenged with SR141716A. As mentioned above, perhaps the intensity of paw tremors and head shakes excluded the possibility of scratching. However, drug interaction between SR141716A and Δ²-THC cannot be ruled out. Because the Δ²-THC suppression of SR141716A induced scratching was not dose-dependent, a drug interaction appears unlikely. High doses of SR141716A elicited a few head shakes and paw tremors, although they were of low intensity and high variability. It is possible that SR141716A produces these effects either by disrupting the normal functioning of the endogenous cannabinoid system or by a direct pharmacological action of its own. The latter seems somewhat unlikely given that the pharmacological effects are identical with those observed during withdrawal.

There has been considerable interest in commonalities between opioids and cannabinoids, particularly with regards to tolerance and dependence. For example, cross-tolerance has been reported by Smith et al. (1994) between Δ⁹-THC and kappa opioid receptor agonists U-50,488H and CI-977. Evidence exists that suggests precipitation of withdrawal by naloxone or drug cessation in rats chronically exposed to Δ⁹-THC elicits behavioral signs suggestive of withdrawal (Hirschhorn and Rosecrans, 1974; Kaymakcalan et al., 1977; Verberne et al., 1980). In addition, Δ⁹-THC has been documented to suppress naloxone-induced precipitated withdrawal in morphine-dependent rats (Bhargava, 1976; Bhargava, 1978; Hine et al., 1975). Conversely, naloxone did not elicit any behavioral effects in monkeys chronically exposed to Δ⁹-THC (Beardsley et al., 1986) nor did SR141716A precipitate morphine withdrawal in morphine-dependent rats (Aceto et al., 1996). Cannabinoids and SR141716A clearly compete for receptor binding sites, whereas there has been no conclusive evidence to suggest that cannabinoids compete with naloxone binding. Therefore, SR141716A appears to be selective for cannabinoids. While it is clear that distinctive endogenous systems are involved in dependence to opioids and cannabinoids, the expression of withdrawal could result from activation of common neurological pathways.

For some classes of drugs, dependence intensity is a function of the degree of tolerance developed. However, Δ⁹-THC is capable of producing a considerable degree of tolerance, yet termination of chronic treatment is not accompanied by a severe withdrawal syndrome. Previous studies in our laboratory have shown pharmacological tolerance in mice after a dose of 10 mg/kg of Δ⁹-THC twice a day for 1 wk (Abood et al., 1993; Fan et al., 1994). It is particularly important to point out that this treatment regimen produces very few overt behavioral effects. The data from this study suggest that using the same treatment regimen, physical dependence developed in addition to pharmacological tolerance. The data from this study as well as from other studies suggests that cannabinoid tolerance (Compton et al., 1990; Pertwee, 1991) and dependence could possibly develop as quickly as 2 days. However, parallel time course studies have not been conducted to establish the relationship between these two phenomena.

Before the discovery of SR141716A, dependence studies had to rely on abrupt cessation of chronic drug administration. This approach led to conflicting data that were difficult to interpret, especially in humans. Common withdrawal signs noted by several investigators were hyperirritability, tremors, sweating, dysphoria, anxiety, negativism, weight loss (decreased appetite) and insomnia (Cohen et al., 1976; Fraser, 1949; Georgotas and Zeidenberg, 1979; Greenberg et al., 1976; Jones and Benowitz, 1976, 1981; Mendelson et al., 1976; Souef, 1967). A diminution of withdrawal signs was observed upon re-administration of Δ⁹-THC (Jones, 1983). However, a major confound with human studies was the anticipation of drug withdrawal.

The availability of SR141716A made it possible to carry out studies to either refuse or reinforce the theory of physical dependence development to cannabinoids. Therefore, it was vital that precipitated withdrawal studies with cannabinoids be conducted. The data from the present investigation together with the previous dependence studies in rats (Aceto et al., 1995, 1996; Tsou et al., 1995) clearly demonstrated the development of cannabinoid dependence.

We were able to develop and characterize a mouse model for cannabinoid dependence. SR141716A induced a precipitated withdrawal syndrome that included involuntary paw tremors and head shakes, disorganized random movements and decrease in grooming and scratching behavior. Paw tremors, head shakes and grooming behavior were dependent on the dose of SR141716A. Moreover, this precipitated withdrawal syndrome could be precipitated in mice treated for only 2 days with Δ⁹-THC.

In summary, in mice as well as in humans (Jones, 1983), the intensity of the dependence depended on the length of treatment time and dose of Δ⁹-THC. It is important to keep in mind that the chronic Δ²-THC regimen used in the mouse model mimics heavy marijuana use. Because the frequency, quantity and duration of drug use dictate the intensity of dependence, it seems unlikely that infrequent marijuana use will result in dependence.

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References


