Introduction

A preponderance of experimental data obtained in animals strongly supports the notion that the reinforcing properties of commonly abused drugs are mediated through the mesolimbic–mesocortical dopamine system originating in the midbrain ventral tegmental area (VTA).\textsuperscript{1,2} Marijuana, a highly abused illicit substance would be suspected, therefore, to alter dopamine activity in reward relevant circuits in the brain, in particular the mesolimbic pathway from the VTA to the nucleus accumbens. Prior studies have shown that the psychoactive constituent of marijuana, \textit{D}9-\textit{tetrahydrocannabinol} (\textit{D}9-\textit{THC}), increases extracellular levels of dopamine in the accumbens and striatum, and induces amphetamine-like ipsiversive turning in rats with unilateral nigrostriatal lesions.\textsuperscript{3–6} Thus, it has been suggested that \textit{D}9-\textit{THC} acts through a presynaptic site of action possibly in the manner of a dopamine reuptake inhibitor. An alternative mechanism by which \textit{D}9-\textit{THC} could augment dopamine neurotransmission has not been fully explored, namely \textit{D}9-\textit{THC}-induced changes in dopamine cell firing. Therefore, the present study was designed to determine the effects of systemic administration of \textit{D}9-\textit{THC} and the non-psychoactive cannabidiol (CBD) on the activity of single dopamine neurons within the VTA and substantia nigra pars compacta (SNC). The effects of the synthetic cannabimimetic aminoalkylindole \textit{WIN} 55,212-2 and its inactive enantiomer \textit{WIN} 55,212-3 were also assessed and compared with those of the natural cannabinoids.

Materials and Methods

Adult male Sprague–Dawley rats weighing 250–350 g were used in all experiments. All animals were housed under a light-dark schedule (07.00–19.00 h) with constant room temperature and free access to food and water. All experimental and surgical manipulations were carried out in accordance with a University of Arizona IACUC approved protocol. Chloral hydrate (350 mg kg\textsuperscript{-1}, i.p.) was used for the induction and maintenance of anesthesia throughout the recording period. The preparation of the animal for i.v. drug injections and a detailed description of the electrophysiological techniques for recording from VTA and SNC dopamine neurons have been described elsewhere.\textsuperscript{7} Inter-spike interval histograms for computing burst activity were constructed off-line from 500 consecutive spikes preceding the onset of the next injection. The criteria used for the definition of bursting parameters have been detailed elsewhere.\textsuperscript{7} VTA and SNC recording sites were made at the following coordinates relative to bregma: posterior 5–5.5 mm, and lateral 0.5–1.0 (VTA) and 1.2–2.0 mm (SNC).

All drugs were injected i.v., and injections were separated by intervals sufficient to collect at least 500 spikes, which in most cases was 2–4 min. Drugs were administered in a cumulative dosing paradigm according to the following protocol: \textit{D}9-\textit{THC} and CBD: 0.125, 0.25, 0.5, 1, 2 and 4 mg kg\textsuperscript{-1}, for a total cumulative dose of 7.875 mg kg\textsuperscript{-1}, and \textit{WIN} 55212-2 and -3: 0.0125, 0.025, 0.05, 0.1, 0.2 and 0.4 mg kg\textsuperscript{-1}, for a total cumulative dose of 0.7875 mg kg\textsuperscript{-1}. Only...
Systemic injections of Δ9-THC produced significant \( p < 0.01 \) dose-dependent increases in dopamine cell firing in both VTA and SNC with maximum increases of 55% in the VTA and 28% in the SNC (Fig. 1, top). An examination of the dose–response curves further reveals that VTA neurons required about one-sixth the dose of Δ9-THC to produce a level of excitation (20%) comparable with that seen in the SNC. These effects appear to be specific to the psychoactive cannabinoid since the non-psychoactive CBD failed to affect these parameters of dopamine cell excitability, even at higher doses (data not shown). The vehicle of DMSO/Tween-80/water was ineffective, producing a maximum change in rate of only –7%. Cannabinoid-induced changes in the number of action potentials contained in bursts were also dose-dependently increased in the VTA from a basal level of 9.7% to 34.5% (Fig. 1, bottom). Bursting in the SNC was virtually unchanged, however, from a baseline of 9.6% to 10.3% (Fig. 2).

The electrophysiological response of dopamine neurons to the cannabimimetic enantioselective WIN 55,212-2 was a significant \( p < 0.01 \) by one-way ANOVA excitation with firing rates in the VTA and SNC increasing by 61.5% and 58.5%, respectively (Fig. 2). Burst firing also increased in both regions from 6.6% to 22.5% in the SNC and from 21% to 31% in the VTA. Again, SNC neurons were less sensitive than those in the VTA, requiring about 2.5 times more WIN 55,212-2 to produce a comparable change (40%) in rate. These effects also appear to be specific since the inactive isomer WIN 55,212-3 failed to change VTA activity; SNC neurons were not tested with the inactive enantiomer.
Discussion

The present results show that Δ9-THC, the psychoactive ingredient in marijuana, can excite midbrain dopamine neurons, in particular those in the VTA which comprise the mesolimbic-mesocortical pathways. Since these dopaminergic circuits are known to play a pivotal role in mediating the reinforcing effects of most drugs of abuse, the increased dopamine drive elicited by the cannabinoids could underlie the abuse property of marijuana.

These data also provide an alternative explanation for the observed increases in extracellular levels of dopamine following the administration of cannabinoids. It had been speculated that cannabinoid-induced increases in dopamine release were mediated through a presynaptic mechanism akin to that seen with dopamine reuptake blockers such as nomifensine. Dopamine reuptake inhibitors inhibit VTA and SNC firing, however. Therefore, the increased incidence of firing and bursting observed in the present study could provide an alternative mechanism by which cannabinoids increase extracellular dopamine levels. Burst activity in particular has been shown to markedly augment transmitter release. Furthermore, the 30% increase in VTA dopamine firing observed here at 1 mg kg\(^{-1}\) would appear to correspond to the 20% increase in dopamine release measured in the nucleus accumbens following the same dose of THC. The differences in the magnitude of change may best be explained by the i.v. vs i.p. routes of administration used in these two studies. Also, the doses leading to dopamine neuronal stimulation and mesolimbic dopamine release are relevant to human cannabinoid pharmacology. Nevertheless, it still remains to be determined whether the euphoria, dysphoria, or even psychotic-like symptoms that have been associated with marijuana use are the result of cannabinoid-evoked activation of dopaminergic mesolimbic-mesocortical pathways.

In some respects the cannabinoid effects reported here bear a resemblance to the biochemical and electrophysiological changes observed in VTA dopamine neurons following the hallucinogen phencyclidine (PCP). Unlike PCP, however, psychoactive cannabinoids do not involve a site of action within the N-methyl-D-aspartate-ion channel complex found on VTA dopamine neurons. Furthermore, since cannabinoid receptors do not appear to reside on dopamine cell bodies, the effects of Δ9-THC and WIN 55,212-2 would be more likely to occur through an alteration of transmitter(s) afferent to the VTA and SNC dopamine neurons. One possibility would be cannabinoid-induced inhibition of local circuit γ-aminobutyric acid neurons within the VTA leading to a disinhibition of dopamine cell firing. However, the finding that injections of Δ9-THC directly into the VTA fails to produce any extracellular dopamine changes in the nucleus accumbens argues against this mechanism. This might also rule out a direct stimulatory effect of cannabinoids on dopamine neurons. Ongoing studies in midbrain slice preparations will help determine whether the cannabinoid-induced excitations result from direct actions on dopamine neurons, local circuit neurons, or other VTA afferents.

The present findings that psychoactive cannabinoids stimulate midbrain dopamine neurons are also of interest given the fact that there appear to be few cannabinoid binding sites in either VTA or SNC. Nevertheless, an examination of autoradiograms of cannabinoid receptor binding of \[^{3}H\]CP-55940 clearly shows labeling of cells in the VTA. Furthermore, cells displaying low levels of hybridization to the mRNA for the cannabinoid receptor have been visualized throughout the substantia nigra and VTA. In addition, there are areas in the CNS where there is a mismatch between cannabinoid receptor density and the extent of metabolic effects of cannabinoids as revealed through 2-deoxyglucose autoradiography.

A surprising result of the present study was the apparent lack of sensitivity of dopamine neurons in the SNC compared to those in the VTA. It is well-known that cannabinoids can produce marked catalepsy presumably by alterations of nigro-striatal dopamine release. The present data, however, would lead to the conclusion that this behavioral effect is likely not mediated through cannabinoid-induced stimulation of SNC neurons, since firing rates increased less than 30%, burst firing only 7% above baseline levels, with maximum effects on firing occurring at an i.v. dose of 8 mg kg\(^{-1}\). This dose of Δ9-THC is considerably greater than that required to elicit catalepsy. Others have also reported that limbic structures are more sensitive than the striatum to Δ9-THC-induced increases in dopamine metabolism. Also, the discriminative stimulus effects of Δ9-THC occur at doses five-fold less than those producing catalepsy. Whether the neurobiological underpinnings for cannabinoid drug discrimination and reinforcement share a common mesolimbic dopamine pathway is unclear. This differential sensitivity of midbrain dopamine neurons is not unique to Δ9-THC, as other reinforcing drugs also produce larger changes in firing rate in VTA than in the SNC.

It is generally accepted that the synthetic aminoalkylindole compound WIN 55,212-2 acts at the same receptor site as the natural cannabinoid moieties. In a battery of behavioral tests this drug produces the classic behavioral changes shown to occur following the administration of Δ9-THC. In the present
study, WIN 55,212-2 produced a similar magnitude of firing of VTA dopamine neurons to that seen with Δ9-THC but at doses generally 10-fold less. As with Δ9-THC, the increased firing rates were accompanied by increases in burst firing. Therefore, one might predict that WIN 55,212-2 would produce effects on dopamine release comparable to Δ9-THC. Notably, SNC neurons seemed more responsive to WIN than to Δ9-THC, and at the highest doses tested WIN 55,212-2 did not attenuate the increased firing rate as seen with Δ9-THC. These apparent differences between the cannabinoid compounds cannot be readily explained. Nevertheless, the effects of WIN 55,212-2 are considered specific since the inactive enantiomer WIN 55,212-3 failed to stimulate dopamine cell firing.

Conclusion

These data show that behaviorally active cannabinoids markedly increase both the firing rate and bursting activity of midbrain dopamine neurons. This mechanism may account for the increases in extracellular mesolimbic dopamine levels observed in vivo microdialysis experiments. Midbrain dopamine neuronal systems are intimately involved in appetitive behaviors and as such may underlie the learning and execution of goal directed behaviors. Thus, it is conceivable that the abuse potential of marijuana may be subserved by cannabinoid-induced activation of the neuronal elements comprising this mesolimbic system.

References


ACKNOWLEDGEMENT: This work is supported through Grant DA 09025 from the National Institute on Drug Abuse.

Received 30 August 1996; accepted 17 October 1996